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## Nucleosides, Nucleotides and Nucleic Acids

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### Applications of LC/MS and Tandem Mass Spectrometry to the Characterization of Nucleosides in Mixtures

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## APPLICATIONS OF LC/MS AND TANDEM MASS SPECTROMETRY TO THE CHARACTERIZATION OF NUCLEOSIDES IN MIXTURES

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**Abstract:** *Liquid chromatography-mass spectrometry (LC/MS) and tandem mass spectrometry (MS/MS) provide new approaches for structural studies of nucleosides, in the nanogram range, in mixtures. Examples are given of the use of LC/MS for rapid screening of synthesis reaction mixtures, and of MS/MS for the detection and characterization of nucleoside isomers in RNA hydrolysates.*

The structural characterization of nucleosides is a problem common to many fields, including synthetic chemistry (e.g., nucleoside analogs), pharmacology (metabolized products), and biochemistry (natural modification in RNA and DNA; xenobiotic modifications). The structure determination of new or unexpected products is particularly difficult in quantities below  $\sim 10^{-6}$  gm, and becomes acute in complex mixtures. Mass spectrometry has played a valuable role in the characterization of nucleosides and related compounds (reviewed in ref. 1) due mainly to its high sensitivity, the availability of various microscale approaches,<sup>2</sup> and the advantages which accrue from its direct combination with chromatography. In particular, the development of LC/MS and tandem mass spectrometry has provided new approaches to the microscale characterization of nucleosides in mixtures (reviewed in ref. 3).

As an extension of LC/MS- and MS/MS-based methods<sup>1,3,4</sup>, some of which have been applied to structural problems involving natural nucleosides,<sup>5</sup> we comment here on two additional types of analyses: (1) use of LC/MS for rapid screening of synthesis reaction mixtures, and (2) initial characterization of nucleoside isomers by MS/MS, without chromatography, in hydrolysates of nucleic acids.

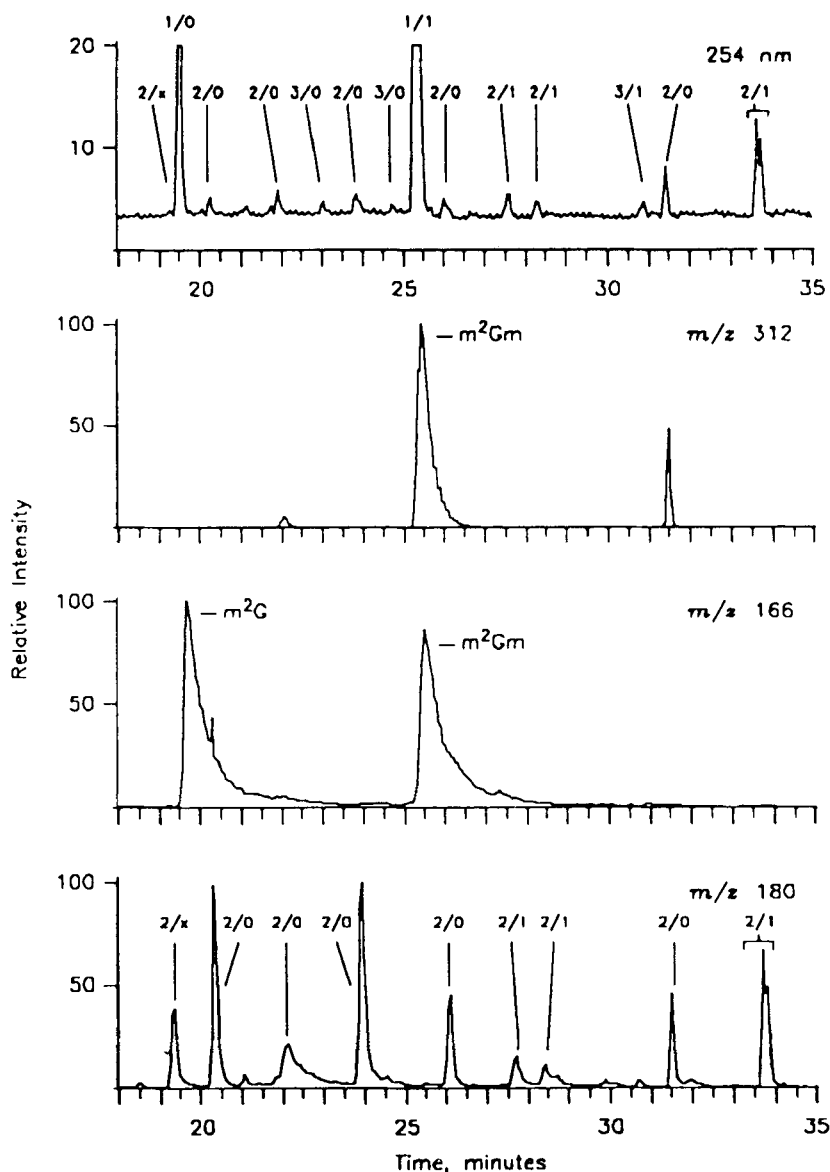


FIG. 1 LC/MS analysis of methylated products of N<sup>2</sup>-methylguanosine (SnCl<sub>2</sub>, CH<sub>2</sub>N<sub>2</sub>/glyme, DMF, 17 hrs, 23°C). Top: UV detection. Peak designations show numbers of methyl groups in base and sugar, respectively, determined from mass spectra; x = unknown number. Bottom panels: examples of mass channels, taken from full mass spectra recorded every 3 sec.; m/z 312, molecular (MH<sup>+</sup>) ion showing overall incorporation of two methyls; m/z 166 and 180, base (BH<sub>2</sub><sup>+</sup>) ions showing incorporation of one and two methyls, respectively, in the base. Instrument: non-commercial quadrupole, conditions previously described.<sup>4</sup>

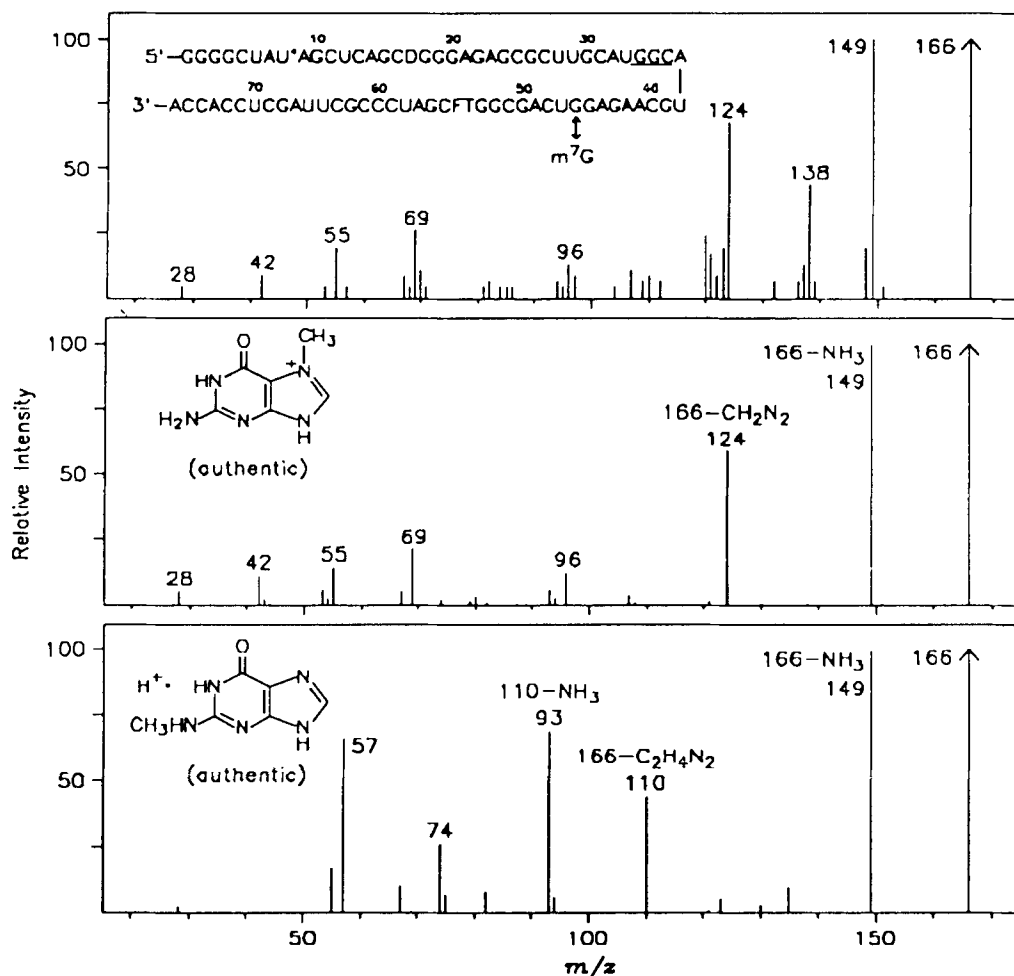


FIG. 2 FAB-MS/MS analysis of nucleosides produced by enzymatic digestion<sup>4</sup> of *E. coli* tRNA<sup>Ala</sup><sub>GCC</sub> corresponding to 5  $\mu$ g of tRNA. Top: product ions from CID of  $m/z$  166 (<25 ng of nucleoside); inset, published sequence.<sup>9</sup> Bottom panels: product ions from CID of  $m/z$  166 ( $BH_2^+$ ) from authentic  $m^7G$  (center) and  $m^2G$  (bottom). Instrument: VG 70-SEQ; dithioerythritol:dithiothreitol (4:1) matrix; Xe FAB gas; Kr collision gas, 30 eV collision energy.

### Screening of Reaction Mixtures by Thermospray LC/MS

Although less sensitive than GC/MS, thermospray LC/MS is applicable to nucleosides in amounts of about 10 ng or greater per HPLC injection (scanned spectra), or 2 ng for selected ion monitoring,<sup>4</sup> and does not require derivatization for enhancement of volatility. With appropriate reversed phase HPLC procedures,<sup>6</sup> these characteristics are advantageous for rapid screening of reaction mixtures when the elution position of the component of interest is not known, either as a guide for subsequent isolation, or for identification of reaction products.

Figure 1 shows a typical example, in which N<sup>2</sup>,2'-O-dimethylguanosine is sought as a product from reaction of diazomethane with N<sup>2</sup>-methylguanosine. The 19.5 min. eluant is readily identified as starting material by its mass spectrum (not shown), and the 25.4 min. eluant as a candidate for the desired product (verified by collection, trimethylsilylation, and EI mass spectrometry<sup>7</sup>). Mass spectrometry provides significant enhancement of signal compared with UV detection (compare UV and m/z 180 channels), with considerable structural information as dictated by choice of mass channels. In appropriate cases an LC column can be omitted, and the reaction solution directly injected.

### Characterization of Nucleoside Isomers in Mixtures by MS/MS

MS/MS can be used, often without the necessity of chromatography, for recognition of modified nucleosides in crude enzymatic digests of RNA.<sup>5,8</sup> Figure 2 shows the detection of 7-methylguanosine in a hydrolysate of isoaccepting tRNA, where it occurs at trace levels at position-46 by partial replacement of guanosine.<sup>9</sup> Its differentiation from other isomers, for example N<sup>2</sup>-methylguanosine, is clearly made by both mass and abundance of ions in the m/z 42-124 range. As a cautionary note, it should be recognized that mixtures of isomers having the same precursor mass, if present, will give overlapping CID spectra whose interpretation may be ambiguous. The MS/MS approach is preferred over LC/MS when nucleotides or other highly polar products, which do not exhibit good thermospray mass spectra, are present in the mixture.

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